

*Hamatocanthoscypha podocarpi*



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***Hamatocanthoscypha podocarpi* Crous, sp. nov.**

*Etymology.* Name refers to the host genus *Podocarpus* from which it was isolated.

*Classification* — *Hamatocanthoscyphaceae*, *Helotiales*, *Leotiomyces*.

*Mycelium* consisting of hyaline, branched, 1.5–2 µm diam hyphae. *Conidiophores* smooth, pale to medium brown, erect, solitary or in clusters, subcylindrical, branched below, 0–4-septate, 12–60 × 3–5 µm. *Conidiogenous cells* 13–40 × 3–4 µm, integrated, terminal and intercalary, subcylindrical, pale brown, smooth, base tapering to long cylindrical, apical venter, 3–9 µm long, slightly flared or not, 2–3 µm diam. *Conidia* in long unbranched chains, aseptate, hyaline, smooth, guttulate, subcylindrical with truncate ends, (6–)7–8(–9) × (1.5–)2 µm.

*Culture characteristics* — Colonies flat, spreading, with folded surface, moderate aerial mycelium and smooth, lobate margin, reaching 20 mm diam after 2 wk at 25 °C. On MEA surface honey, reverse cinnamon. On PDA surface honey with isabelline in outer region, reverse isabelline. On OA surface honey.

*Typus.* SOUTH AFRICA, Western Cape Province, Knysna, on leaf spots of *Podocarpus latifolius* (*Podocarpaceae*), Nov. 2018, *M.J. Wingfield*, HPC 2710 (holotype CBS H-24349, culture ex-type CPC 37055 = CBS 146626; ITS, LSU, *actA* and *rpb2* sequences GenBank MT373365.1, MT373348.1, MT375095.1 and MT375100.1, MycoBank MB835399).

*Notes* — The genus *Chalara* as circumscribed by Nag Raj & Kendrick (1976) is polyphyletic and awaits revision. *Hamatocanthoscypha podocarpi* is phylogenetically allied to the type species of *Hamatocanthoscypha*, *H. laricionis* (Svrček 1977), and placed in this genus based on DNA similarity. Several species of '*Chalara*' have been described from *Podocarpus*, namely *C. brevipes* (conidia (6–)8.9(–12) × 1.5–2 µm), *C. novaezelandiae* (conidia (5–)6.4(–8) × 1–1.5 µm), *C. cylindrosperma* (conidia (5.5–)11(–17) × (1.5–)1.9(–2.5) µm), *C. fusidioides* (conidia (4.5–)7.7(–12) × (1.5–)2.1(–3.5) µm), *C. acuaria* (conidia (12–)16(–20) × (2–)2.7(–3.5) µm) and *C. bicolor* (conidia 7-septate, (45–)50–60(–71) × 5.5–6 µm) (Nag Raj & Kendrick 1975). Of these, *H. podocarpi* is most similar to *C. brevipes*, but can be distinguished in having smaller conidiogenous cells, and conidiophores that are aggregated in clusters.

Based on a megablast search of NCBI's GenBank nucleotide database, the closest hits using the ITS sequence had highest similarity to numerous sequences wrongly labelled as '*Infundichalara microchona*' (e.g., strain KRP75-5, GenBank HM036588.1; Identities = 531/537 (99 %), 2 gaps (0 %)), *Chalara holubovae* (strain CCF 3978, GenBank NR\_154760.1; Identities = 483/501 (96 %), 3 gaps (0 %)), and *Hamatocanthoscypha laricionis* (voucher TNS-F13530, GenBank JN033441.1; Identities = 540/567 (95 %), 4 gaps (0 %)). Closest hits using the LSU sequence are *Leptodontidium beauverioides* (strain CBS 672.76, GenBank MH872794.1; Identities = 836/840 (99 %), no gaps), *Tricladium caudatum* (strain CCM F-13498, GenBank GQ477318.1; Identities = 833/837 (99 %), no gaps), and *Chalara constricta* (strain CBS 248.76, GenBank FJ176256.1; Identities = 825/829 (99 %), no gaps). No significant hits were obtained when the *actA* and *rpb2* sequences were used in blastn and megablast searches.

*Colour illustrations.* Walkway in the Knysna forest. Conidiophores with conidiogenous cells; conidia. Scale bars = 10 µm.

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