Perenniporia brasiliensis
**Perenniporia brasiliensis** Lira, A.M.S. Soares, Ryvarden & Gibertoni, *sp. nov.*

**Etymology.** Referring to the country where this fungus was collected, Brazil.

Classification — *Polyporaceae, Polyporales, Agaricomycetes.*

**Basidiomata** annual, resupinate, smooth and even, hard to brittle, 10 × 1.5 cm in the holotype and 0.5 mm thick; pore surface cream greyish to tan (31 vinaceous buff to 10 cinnamon); pores slightly thick-walled, round to angular, mostly 6–7 per mm; dissepiments entire, thick; tubes concolorous with the pore surface, up to 0.5 mm deep; context about 100 mm thick, cottony and concolorous with the pore surface; margin smooth, narrow and concolorous with the pore surface. *Hyphal system* dimitic, generative hyphae thin-walled, smooth and with clamps, 2–4 μm wide, skeletal hyphae weakly dextrinoid, 2–3 μm. *Cystidia* or other sterile elements absent. *Basidia* 14–20 × 4–6 μm, clavate with four sterigmata. *Basidiospores* 3–4 × 2–4 μm, globose to subglobe, hyaline, thick-walled and dextrinoid.


Notes — Based on a BLASTn search of NCBI’s GenBank database, the closest hits using the ITS sequence are *Perenniporia* sp. (GenBank KT156689; Identities = 588/598 (98 %), Gaps = 1/598 (0 %)), *Dichomitus squalen* (GenBank KM411455; Identities = 631/666 (95 %), Gaps = 4/666 (0 %)), and *P. tenuis* (GenBank JQ001859: Identities = 631/667 (95 %), Gaps = 7/667 (0 %)). Using the LSU sequence, the highest similarity was to *P. aridula* (GenBank JQ001847; Identities = 801/817 (98 %), Gaps = 7/817 (0 %)), *P. aridula* (GenBank JQ001846; Identities = 801/817 (98 %), Gaps = 7/817 (0 %)), and *P. tibetica* (Gen-Bank JF706332; Identities = 801/817 (98 %), Gaps = 7/817 (0 %)). Although genetically close to *P. aridula*, *P. tenuis* and *P. tibetica, P. brasiliensis* is morphologically different (Table 1 - see FP 612). *Perenniporia brasiliensis* is similar to *P. albo-incarnata, P. centrali-africana*, and *P. guyanensis*, sharing the same whitish colour. However, they are micro-morphologically different (Table 1 - see FP 612).

Carla R.S. Lira, Adriene M. Soares, Tatiana B. Gibertoni, Departamento de Micologia Prof. Chaves Batista, Universidade Federal de Pernambuco, Recife, Brazil; e-mail: carla-rejane@hotmail.com, adrienemsoares@gmail.com & tbgibertoni@hotmail.com

Leif Ryvarden, University of Oslo, Institute of Biological Sciences, P.O. Box 1066, Blindern, N-0316, Oslo, Norway; e-mail: leif.ryvarden@ibv.uio.no

Cony Decock, Mycotheca de l’Université catholique de Louvain (MUCL, BCCMTM), Earth and Life Institute – Microbiology (ELIM), Université catholique de Louvain, Croix du Sud 2 bte L7.05.06, B-1348, Louvain-la-Neuve, Belgium; e-mail: cony.decock@uclouvain.be